## **PCT**

#### INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference FOR FURTHER see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.							
International application No.	International filing date (day/mont	h/year) (Earliest) F	Priority Date (day/month/year)				
PCT/GB 99/02652	12/08/1999		13/08/1998				
Applicant							
ZENECA LIMITED et al.							
This International Search Report has bee according to Article 18. A copy is being to			ransmitted to the applicant				
This International Search Report consists  X It is also accompanied by	of a total of sh a copy of each prior art document o						
Basis of the report							
a. With regard to the language, the language in which it was filed, un	international search was carried out less otherwise indicated under this i		rnational application in the				
the international search w Authority (Rule 23.1(b)).	as carried out on the basis of a tran	slation of the internation	al application furnished to this				
b. With regard to any <b>nucleotide ar</b> was carried out on the basis of th	•	ed in the international ap	pplication, the international search				
	onal application in written form.	adabla form					
	ernational application in computer re	adable form.					
	furnished subsequently to this Authority in written form.						
the statement that the sul	o this Authority in computer readble bsequently furnished written sequen		eyond the disclosure in the				
l	is filed has been furnished. Ormation recorded in computer read	able form is identical to t	the written sequence listing has been				
furnished	·						
2. Certain claims were fou	nd unsearchable (See Box I).						
3. Unity of invention is lac	king (see Box II).		•				
4. With regard to the title,							
X the text is approved as su	ibmitted by the applicant.						
the text has been established by this Authority to read as follows:							
5. With regard to the abstract,							
X the text is approved as su	• • • • • • • • • • • • • • • • • • • •						
	the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the date of mailing of this international search report, submit comments to this Authority.						
6. The figure of the <b>drawings</b> to be published with the abstract is Figure No.							
as suggested by the appl	icant.		None of the figures.				
because the applicant fail	led to suggest a figure.						
because this figure better	characterizes the invention.						

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/31 C12N15/82 A01H5/00

According to International Patent Classification (IPC) or to both national classification and IPC

#### B. FIELDS SEARCHED

 $\frac{\text{Minimum documentation searched (classification system followed by classification symbols)}}{IPC-7-C07K-C12N-A01H}$ 

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

		Relevant to claim No.
Category °	Citation of document, with indication, where appropriate, of the relevant passages	nelevant to claim No.
Α	DE 195 48 301 C (DUERING KLAUS DR) 27 February 1997 (1997-02-27) the whole document	1-6
Α	WO 97 48719 A (BECKERMAN JANNA L ;TEXAS A & M UNIVERSITY SYST (US); ZHANG LEI (US) 24 December 1997 (1997-12-24) the whole document	1-6
A	WO 96 29392 A (UNISEARCH LTD ;KJELLEBERG STAFFAN (AU); STEINBERG PETER (AU); NYS) 26 September 1996 (1996-09-26) the whole document 	2

X Further documents are listed in the continuation of box C.	Patent family members are listed in annex.
<ul> <li>Special categories of cited documents:</li> <li>"A" document defining the general state of the art which is not considered to be of particular relevance</li> <li>"E" earlier document but published on or after the international filing date</li> <li>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</li> <li>"O" document referring to an oral disclosure, use, exhibition or other means</li> <li>"P" document published prior to the international filing date but later than the priority date claimed</li> </ul>	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention  "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone  "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combined with one or more other such documents, such combination being obvious to a person skilled in the art.  "&" document member of the same patent family
Date of the actual completion of the international search  2 November 1999	Date of mailing of the international search report $17/11/1999$
Name and mailing address of the ISA  European Patent Office, P.B. 5818 Patentlaan 2  NL - 2280 HV Rijswijk  Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  Fax: (+31-70) 340-3016	Authorized officer  Kania, T

	<u></u>		
C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT	Relevant to claim No.		
Category Citation of document, with indication, where appropriate, of the relevant passages	Helevant to claim No.		
GRAY K. ET AL.: "Cell-to cell signaling in the symbiotic nitrogen-fixing bacterium Rhizobium leguminosarum: autoinduction of a stationary phase and rhizosphere-expressed genes" JOURNAL OF BACTERIOLOGY, vol. 178, no. 2, 1996, pages 372-376, XP002084285 the whole document	3		
ROSEMEYER ET AL: "luxI- and luxR-homologous genes of Rhizobium etli CNPAF512 contribute to synthesis of autoinducer molecules and nodulation of Phaseolus vulgaris"  JOURNAL OF BACTERIOLOGY, vol. 180, no. 4, 1 February 1998 (1998-02-01), pages 815-821, XP002084284 ISSN: 0021-9193	3		
THROUP J. ET AL.: MOLECULAR MICROBIOLOGY, vol. 17, 1996, pages 345-56, XP002121181 cited in the application the whole document	1-6		
ROBSON N D ET AL: "Bacterial N-acyl-homoserine-lactone-dependent signalling and its potential biotechnological applications" TRENDS IN BIOTECHNOLOGY, vol. 15, no. 11, 1 November 1997 (1997-11-01), page 458-464 XP004092668 ISSN: 0167-7799 see the whole document; esp. p.461 r. col.	1-6		
SWIFT S ET AL: "Quorum sensing: a population-density component in the determination of bacterial phenotype" TIBS TRENDS IN BIOCHEMICAL SCIENCES, vol. 21, no. 6, 1 June 1996 (1996-06-01), page 214-219 XP004050894  ISSN: 0968-0004 cited in the application the whole document	1-6		

1

mation on patent family members

ĺ	rnational Application No	
	PCT/GB 99/02652	

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
DE 19548301 C	27-02-1997	NONE	
WO 9748719 A	24-12-1997	AU 3570997 A	07-01-1998
WO 9629392 A	26-09-1996	AU 708962 B	19-08-1999
		AU 4999696 A	08-10-1996
		BR 9607661 A	16-06-1998
		CA 2215797 A	26-09-1996
		CN 1185173 A	17-06-1998
		EP 0815201 A	07-01-1998
		JP 11502108 T	23-02-1999
		NZ 303630 A	26-01-1998



## **PCT**

## INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

• •		t's file reference	FOR FURTHER ACTION	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)
PPD 5036	0/WC	)		
mental spring			International filing date (day/month	1
PCT/GB99			12/08/1999	13/08/1998
International C12N15/3		t Classification (IPC) or na	tional classification and IPC	
Applicant				
ZENECA	LIMI	ΓED et al.		
1. This in and is	terna trans	tional preliminary exam mitted to the applicant a	ination report has been prepare according to Article 36.	d by this International Preliminary Examining Authority
2. This R	EPOI	RT consists of a total of	10 sheets, including this cover	sheet.
be (s	een ai ee Ru	mended and are the bar	sis for this report and/or sheets 07 of the Administrative Instruct	ne description, claims and/or drawings which have containing rectifications made before this Authority ions under the PCT).
3. This re	_		ating to the following items:	
1		Basis of the report		
		Priority	opinion with regard to novelty in	nventive step and industrial applicability
111	⋈			toniaro stop and manatim approach,
V	×	Reasoned statement	under Article 35(2) with regard to ions suporting such statement	o novelty, inventive step or industrial applicability;
VI		Certain documents ci		
VII	$\boxtimes$	Certain defects in the	international application	
VIII	Ø	Certain observations of	on the international application	
Date of sub	omissio	on of the demand	Date o	of completion of this report
15/12/19	99		13.11	2000
Name and preliminary	exam	g address of the internation ining authority:	nal Autho	rized officer
<u></u>	D-8	opean Patent Office 0298 Munich . +49 89 2399 - 0 Tx: 5236	Surd	ej; P
I ——		· +49 89 2399 - 4465		hone No. ±49.89.2399.7334

## INTERNATIONAL PRELIMINARY EXAMINATION REPORT

I. Basis of th r port

International application No. PCT/GB99/02652

1.	response to an invitation	lrawn on the basis of (substitute sheets which have been fumished to the receiving Office in on under Article 14 are referred to in this report as "originally filed" and are not annexed to lo not contain amendments (Rules 70.16 and 70.17).):		
	1-12	as originally filed		
Claims, No.:				
	1-6	as originally filed		
Drawings, sheets:				
	1/1	as originally filed		
2.	With regard to the lan	guage, all the elements marked above were available or furnished to this Authority in the		

language in which the international application was filed, unless otherwise indicated under this item.

These elements were available or furnished to this Authority in the following language: , which is:

		the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
		the language of publication of the international application (under Rule 48.3(b)).
		the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).
3.	Wit	h regard to any <b>nucleotide and/or amino acid sequence</b> disclosed in the international application, the mational preliminary examination was carried out on the basis of the sequence listing:
		contained in the international application in written form.
		filed together with the international application in computer readable form.
		furnished subsequently to this Authority in written form.
		furnished subsequently to this Authority in computer readable form.
		The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

☐ The statement that the information recorded in computer readable form is identical to the written sequence

listing has been furnished.

4. The amendments have resulted in the cancellation of:

☐ the description, pages:
☐ the claims, Nos.:

# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/GB99/02652

		the drawings,	sheets:			
5.	☐ This report has been established as if (some of) the amendments had not been made, since they have be considered to go beyond the disclosure as filed (Rule 70.2(c)):					
		(Any replacement sh report.)	eet contain	ing such i	amendments must be referred to under item 1 and annexed to this	
6.	Add	litional observations, i	f necessary	<b>'</b> :		
IV.	. Lac	ck of unity of invention	on			
				t or pay a	additional fees the applicant has:	
		restricted the claims.				
	Ø	paid additional fees.				
		paid additional fees	under prote	st.		
		neither restricted no	paid additi	onal fees	s.	
2.		This Authority found that the requirement of unity of invention is not complied and chose, according to Rule 68.1, not to invite the applicant to restrict or pay additional fees.				
3.	Thi	This Authority considers that the requirement of unity of invention in accordance with Rules 13.1, 13.2 and 13.3				
	□ complied with.					
	Ø	not complied with for see separate sheet		ng reasor	ons:	
4.		nsequently, the follow amination in establishi			mational application were the subject of international preliminary	
	×	all parts.				
		the parts relating to	claims Nos			
V	. Re	asoned statement u ations and explanati	nder Articlo ons suppo	e 35(2) w rting suc	with regard to novelty, inventive step or industrial applicability; sch statement	
1	. Sta	atement				
	No	velty (N)	Yes: No:	Claims Claims		
	Inv	ventive step (IS)	Yes:	Claims		

# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No. PCT/GB99/02652

Industrial applicability (IA)

Yes:

Claims 1-6

No: Claims

2. Citations and explanations see separate sheet

## VII. Certain defects in the international application

The following defects in the form or contents of the international application have been noted: see separate sheet

## VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made: see separate sheet

Reference is made to the following documents:

D1: DE 195 48 301 C (DUERING KLAUS DR) 27 February 1997 (1997-02-27)

D2: WO 97 48719 A (BECKERMAN JANNA L ;TEXAS A & M UNIVERSITY SYST (US); ZHANG LEI (US) 24 December 1997 (1997-12-24)

D3: GRAY K. ET AL.: 'Cell-to cell signaling in the symbiotic nitrogen-fixing bacterium Rhizobium leguminosarum: autoinduction of a stationary phase and rhizosphere-expressed genes' JOURNAL OF BACTERIOLOGY, vol. 178, no. 2, 1996, pages 372-376

D4: ROSEMEYER ET AL: 'luxl- and luxR-homologous genes of Rhizobium etli CNPAF512 contribute to synthesis of autoinducer molecules and nodulation of Phaseolus vulgaris' JOURNAL OF BACTERIOLOGY, vol. 180, no. 4, 1 February 1998 (1998-02-01), pages 815-821

D5: THROUP J. ET AL.: MOLECULAR MICROBIOLOGY, vol. 17, 1996, pages 345-56, cited in the application

D6: ROBSON N D ET AL: 'Bacterial N-acyl-homoserine-lactone-dependent signalling and its potential biotechnological applications' TRENDS IN BIOTECHNOLOGY, vol. 15, no. 11, 1 November 1997 (1997-11-01), page 458-464

D7: SWIFT S ET AL: 'Quorum sensing: a population-density component in the determination of bacterial phenotype' TIBS TRENDS IN BIOCHEMICAL SCIENCES, vol. 21, no. 6, 1 June 1996 (1996-06-01), page 214-219, cited in the application

D8: WOOD DW AND PIERSON LS: 'The phzl gene of Pseudomonas aureofaciens 30-84 is responsible for the production of a diffusible signal required for phenazine antibiotic production' GENE, vol. 168, no. 1, 2 February 1996 (1996-02-02), page 49-53. This document is not cited in the International Search Report but it is known to the applicant (page 6, line 17 of the description).

#### Introduction

The application discloses methods for the protection of plants against bacterial infection and/or virus infection transmitted by bacteria, methods of enhancing interaction between a rhizobacterium and a plant and a recombinant plant genome carrying a gene to produce the bacterial pheromone N-acyl-L-

**EXAMINATION REPORT - SEPARATE SHEET** 

homoserine lactone in plants.

#### Re Item IV

#### Lack of unity of invention

- 1. The application discloses an alternative method to protect plants against pathogens using the expression of bacterial pheromone N-acyl-L-homoserine lactones (or analogues thereof) in the said plant.

  Genes required for the expression of N-acyl-L-homoserine lactones are known from the prior art (D1, D3-D8), for example all the genes referred to in claim 4 are known. D2 discloses transgenic plants producing fungal pheromone (or pheromone analogues; from page 23, line 26 to page 26, line 19) to confer resistance to fungal infection to the said plant (for example, claims 1-2, page 5, lines 24-26; page 57, lines 21-32; page 58, lines 1-19).
- The application contains independent claims which refer to two different technical 2. problems. Claims 1-2 refer to a method for the protection of plants against bacterial infection and/or virus infection transmitted by bacteria and claim 3 refers to a method of enhancing interaction between a rhizobacterium and a plant. The same solution is provided to solve the two different technical problems, namely introducing into the genome of the plant by transformation the ability to synthesise N-acyl-L-homoserine lactone. However, the two different problems are not linked to each other by a special (new and inventive) technical feature in the sense of Rule 13.2 PCT since the production of transgenic plants with known genes required for the expression of the bacterial pheromone N-acyl-L-homoserine lactone (see point 1) is obvious considering D2 in combination with any of the documents D1, D3-D8 at the priority date of the application. (Thus, it is considered that claim 5 lacks an inventive step and does not provide an inventive link). Therefore, the said problems are not so linked as to form a single general inventive concept as required by Rule 13.1 PCT.
- 3. 2 separate inventions are therefore defined:
  - 1. Claims 1-2 (completely) and 4-6 (partially): Method of protecting a plant against bacterial infection and/or virus infection transmitted by bacteria using transgenic plants producing N-acyl-L-homoserine lactones (or analogues thereof).

- 2. Claims 3 (completely) and 4-6 (partially): Method of enhancing interaction between a rhizobacterium and a plant using transgenic plants producing N-acyl-Lhomoserine lactones.
- In response to the invitation mailed on 17 May 2000, the applicant decided to pay 4. additional fees for the invention 2 identified by the International Preliminary Examination Authority. Therefore, the International Preliminary Examination Report is established on the entire application as filed.

#### Re Item V

Reasoned statement under Article 35(2) PCT with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

Novelty and inventive step (Art. 33(1) - (3) PCT)

Invention 1: Claims 1-2 (completely) and 4-6 (partially).

Claims 1-2 and 4-6 are new since they refer to a method and recombinant plant 5. genome to protect plants against pathogen infection by the expression in planta of acylated homoserine lactones which appear not to be disclosed in the prior art. However, the said claims do not involve an inventive step in the light of document D1 in combination with D2. Considering the said documents, the technical problem of invention 1 is to provide an alternative way to protect plant against pathogen infection.

The closest prior art D1 discloses a method to protect plants against pathogens (such as Erwinia corotovora, column 1, line 36) by inhibiting the pheromone acylated homoserine lactone regulated processes in microorganisms using the expression in planta of antibodies against acylated homoserine lactone. D2 discloses transgenic plants producing fungal pheromone to confer resistance to fungal infection to the said plant (page 5, 5th paragraph; page 57, lines 21-32; page 58, lines 1-19).

The skilled person faced with the technical problem of D1 would combine the solution disclosed in D2, because D2 discloses clearly the use of plant

transformed with genes producing pheromone to protect plants against pathogenic infection; thus, the person skilled in the art would arrive at the claimed subject-matter. Therefore, using the expression of bacterial pheromone in transgenic plant to protect the said plant against bacterial infection seems to be obvious considering D1 and D2 together and it cannot be seen where the inventive step lies in Claims 1 and 3-6.

From D4-D7, it is clear that the bacterial pheromone can be selected from the group consisting of the list of genes given in claim 4.

Claim 2 is not inventive in the light of D2 and any of D3, D6 and D7. A method to 6. protect plants against pathogens by the expression in planta of an analogue of acylated homoserine lactone is referred to in claim 2 of the application. D3 discloses furanone compounds (for example compound 4, page 9, paragraphs 2 and 3) to inhibit acylated homoserine lactone mediated processes in pathogen such as Erwinia corotovora and Pseudomonas aeruginosa. D7 discloses also analogues of acylated homoserine lactone and their role in the inhibition of the signalling pathway (from page 461, 2nd column, last paragraph to page 462, 2nd column, 1st paragraph). Analogues of acylated homoserine lactone are also mentioned in D6 (page 215, 1st column, 2nd paragraph and 2nd column, 1st paragraph; page 218). D2 discloses also analogues of pheromone (page 4, line 4; pages 23-29) and their use in inhibition of pheromone mediated processes in pathogen. The combination of D2 with any of D3, D6 or D7 is obvious since the person skilled in the art would have contemplate the introduction into the genome of plant by transformation of the ability to synthesise analogues of acylated homoserine lactone instead of acylated homoserine lactone. Consequently, no inventive step is acknowledged for claim 2.

Invention 2: Claims 3 (completely) and 4-6 (partially).

7. Claims 3 and 4-6 are new since they refer to method and recombinant plant genome to enhance interaction between a rhizobacterium and a plant by the expression *in planta* of acylated homoserine lactones which appear not to be disclosed in the prior art. However, the said claims do not involve an inventive step in the light of document D8 in combination with D2. The technical problem of invention 2 is to provide an alternative way to enhance interaction between a

rhizobacterium and a plant.

The closest prior art document D8 discloses that the pheromone phzl of Pseudomonas aureofaciens is required for the production of the antibiotic phenazine (e.g. page 51, left column, last paragraph and following page). D8 also discloses that the phenazine protects plants against pathogens (see e.g. page 49, introduction) and exogenously-provided acylated homoserine lactone is capable of restoring phenazine production to the disarmed Pseudomonas aureofaciens phzl strain. D2 discloses transgenic plants producing fungal pheromone to confer resistance to fungal infection to the said plant (page 5, 5th paragraph; page 57, lines 21-32; page 58, lines 1-19).

The skilled person faced with the technical problem of the application would combine the teaching of D8 with the solution disclosed in D2, because D2 discloses clearly the use of plant transformed with pheromone genes to produce pheromone in the rhizosphere and D8 discloses the enhanced plant protection by rhizobacteria which is conferred by the acylated homoserine lactone production. Therefore, using the expression of bacterial pheromone in transgenic plant to enhance interaction between a rhizobacterium and a plant seems to be obvious considering D8 and D2 together and it cannot be seen where the inventive step lies in claims 3 and 4-6.

#### Re Item VII

Certain defects in the international application

Contrary to the requirements of Rule 5.1(a)(ii) PCT, the relevant background art 8. disclosed in the documents D1-D4 and D6 is not mentioned in the description, nor are these documents identified therein.

#### Re Item VIII

Certain observations on the international application

The technical problem of the application does not seem to be solved in the 9. application. Although it is shown that the production of N-acyl-L-homoserine lactone by plant can induce a response in bacteria, there is no indication that plants are actually protected by the said production. Thus, the solution is not

sufficiently disclosed and there is a lack of support for claims 1 and 2 (Art. 5 and 6 PCT).

- 10. Claims 1-3 and 5 do not meet the requirements of Article 6 PCT in that the matter for which protection is sought is not clearly defined. The following functional statements do not enable the skilled person to determine which technical features are necessary to perform the stated functions: synthesis of N-acyl-L-homoserine lactone in plant. The claims attempt to define the subject-matter in terms of the result to be achieved. The technical features necessary for achieving this result should be added. Therefore, the said claims lack clarity and the scope of the said claims is not clear (Art. 6 PCT).
- 11. Claim 5 refers to a "recombinant plant genome". A genome is not an entity which can be isolated as such and cannot be claimed as such. Thus, the scope of the claim is not clear (Art. 6 PCT).
- 12. Claim 2 appears not to be supported by the description. Only methods using the production of natural N-acyl-L-homoserine lactones are disclosed in the application and nowhere in the description is an indication of what an analogue of N-acyl-L-homoserine lactone might be. Therefore, there is insufficiency of disclosure and a lack of support in the description for claim 2 and the scope of the claim is not clear (Art. 5 and 6 PCT).
- 13. Part of claim 5 is not supported by the description as required by Article 6 PCT, as its scope is broader than justified by the description. The reasons therefore are the following: in claim 5, it is referred to a recombinant plant genome containing a gene construct for the expression of a "response regulator" of a N-acyl-Lhomoserine lactone. However, the expression of a response regulator is not sufficiently disclosed and it is not supported by the description since no response regulator are shown to be expressed in plant in the application (Art. 5 and 6 PCT).
- 14. In claim 3, the expression "enhancing interaction" is not clear since it cannot be seen what kind of interaction between a rhizobium and a plant is referred to. The scope of the said claim is therefore not clear (Art. 6 PCT).



#### REQUEST

For receiving Office use only	
International Application No.	
International Filing Date	
Name of receiving Office and "PCT International Application"	

The undersigned requests that the present international application be processed according to the Patent Cooperation Treaty. Applicant's or agent's file reference (if desired) (12 characters maximum) PPD 50360/WO Box No. I TITLE OF INVENTION EXPRESSION OF BACTERIAL SIGNAL MOLECULES IN PLANT Box No. II APPLICANT Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.) This person is also inventor. ZENECA Limited 15 Stanhope Gate Telephone No. London W1Y 6LN 01344 414365 United Kingdom Facsimile No. 01344 481112 Teleprinter No. State (that is, country) of nationality State (that is, country) of residence: ĠB This person is applicant all designated all designated States except the United States of America for the purposes of: the United States of America only States the States indicated in the Supplemental Box FURTHER APPLICANT(S) AND/OR (FURTHER) INVENTOR(S) Box No. III Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (that is, country) of residence if no State of residence is indicated below.) This person is: FRAY Rupert George applicant only Division of Plant Science School of Biological Sciences applicant and inventor University of Nottingham Sutton Bonington Campus inventor only (If this check-box is marked, do not fill in below.) Loughborough LE12 5RD GB State (that is, country) of nationality: State (that is, country) of residence: GB This person is applicant all designated all designated States except the United States of America for the purposes of: the United States of America only States the States indicated in the Supplemental Box Further applicants and/or (further) inventors are indicated on a continuation sheet. AGENT OR COMMON REPRESENTATIVE; OR ADDRESS FOR CORRESPONDENCE Box No. IV The person identified below is hereby/has been appointed to act on behalf of the applicant(s) before the competent International Authorities as: agent common representative (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country.) Name and address: Telephone No. **HUSKISSON Frank Mackie** 01344 414822 Intellectual Propery Department ZENECA Agrochemicals Facsimile No. PO Box 3538 01344 481112 Jealott's Hill Research Station, Bracknell RG42 6YA Teleprinter No. United Kingdom Adress for correspondence: Mark this check-box where no agent or common representative is/has been appointed and the space above is used instead to indicate a special address to which correspondence should be sent.

Continuation of Box No. III		2000
If none of the	001 610	183 2000
Name and address: (Family name follo The address must include postal code and Box is the applicant's State (that is, coun	S1 H	Klins Beaclam
THROUP John Peter Smith Kline Beecham Pharmaceuticals R&D UP1345 South Collegeville Ro PO Box 5089		vouple nationality
Collegeville PA 19426 United States of America		_ , / ~ 4
State (that is, country) of nationality:		18199 GB
This person is applicant all de for the purposes of:	signated all designated States except the United States of America	the United States of America only the Supplemental Box
Name and address: (Family name followe The address must include postal code and no Box is the applicant's State (that is, country	d by given name; for a legal entity, full official ame of country. The country of the address ind ) of residence if no State of residence is indica	designation. licated in this ted below.) This person is:
WALLACE Andrew David Unilever Research Colworth Laboratory		applicant only  applicant and inventor
Colworth House Sharnbrook Bedfordshire MK44 1LQ		inventor only (If this check-box is marked, do not fill in below.)
State (that is, country) of nationality:		hat is, country) of residence: GB
This person is applicant all des	signated all designated States except the United States of America	the United States of America only the States indicated in the Supplemental Box
Name and address: (Family name follower The address must include postal code and na Box is the applicant's State (that is, country,	d by given name; for a legal entity, full official une of country. The country of the address indi ) of residence if no State of residence is indical	designation. icated in this ted below.)  This person is:
GRIERSON Donald Division of Plant Science School of Biological Sciences		applicant only
University of Nottingham Sutton Bonington Campus Loughborough LE12 5RD GB		inventor only (If this check-box is marked, do not fill in below.)
State (that is, country) of nationality:	State (th	nat is, country) of residence: GB
This person is applicant all defor the purposes of:	signated all designated States except the United States of America	the United States the States indicated in the Supplemental Box
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STEWART Gordon Sidney And School of Pharmaceutical Scien University of Nottingham University Park	•	applicant only  applicant and inventor
Nottingham GB	·	inventor only (If this check-box is marked, do not fill in below.)
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This person is applicant all des for the purposes of:	signated all designated States except the United States of America	the United States the States indicated in the Supplemental Box
Further applicants and/or (furthe	r) inventors are indicated on another cont	inuation sheet

Box	No.V	DESIGNATIO. F STATES							
The	follow	ing designations are hereby made under Rule 4.9	9(a) (r	nark ti	he applicable check hoves: at least one must be marked.				
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Intellectual Property Department
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Jealott's Hill Research Station PO Box 3538, Bracknell, Berkshire RG42 6YA, United Kingdom

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Sheet No. 5.....

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13 August 1998	9817707.4	GB					
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This international application c the following number of sheet	ontains This interna	tional application is accompan	nied by the item(s) mark	ed below:			
request : 5	1. X fee ca	alculation sheet					
description (excluding	2. 🔲 separ	ate signed power of attorney					
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FEE CALCULATION SHEET	International application No.
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CALCULATION OF PRESCRIBED FEES	
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3. INTERNATIONAL FEE	
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International filing date: 12 August 1999 (12.08.99)	Priority date: 13 August 1998 (13.08.98)
Applicant: FRAY, Rupert, George et al	
The designated Office is hereby notified of its election made in the demand filed with the International preliminar   15 December  in a notice effecting later election filed with the International preliminar   15 December	ry Examining Authority on: 1999 (15.12.99)
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**OF A CHANGE** (PCT Rule 92bis.1 and Administrative Instructions, Section 422) Bracknell, Berkshire RG42 6YA Date of mailing (day/month/year) 18 April 2001 (18.04.01) Applicant's or agent's file reference IMPORTANT NOTIFICATION PPD 50360/WO International application No. International filing date (day/month/year) PCT/GB99/02652 12 August 1999 (12.08.99) 1. The following indications appeared on record concerning: X the applicant the inventor the agent the common representative State of Nationality State of Residence Name and Address GB GB ZENECA LIMITED 15 Stanhope Gate London W1Y 6LN Telephone No. United Kingdom Facsimile No. Teleprinter No. 2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning: X the name the person the address the nationality the residence State of Nationality State of Residence Name and Address GB SYNGENTA LIMITED Fernhurst Telephone No. Haselmere Surrey GU27 3JE United Kingdom Facsimile No. Teleprinter No. 3. Further observations, if necessary: This is only a change of name and address, no transfer of patent or other rights has occureed. The agent's address has also been changed accordingly. 4. A copy of this notification has been sent to: the receiving Office the designated Offices concerned the International Searching Authority the elected Offices concerned the International Preliminary Examining Authority other: Authorized officer

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Date of mailing (day/month/year) 26 July 2001 (26.07.01)	ROYA	AUME-UNI			
Applicant's or agent's file reference PPD 50360/WO International application No. PCT/GB99/02652	IMPORTANT NOTIFICATION  International filing date (day/month/year) 12 August 1999 (12.08.99)				
The following indications appeared on record concerning:      X the applicant the inventor	the agen	t the commo	on representative		
Name and Address SYNGENTA LIMITED		State of Nationality  GB	State of Residence GB		
Fernhurst Haselmere Surrey GU27 3JE		Telephone No.			
United Kingdom		Facsimile No.			
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(71) Applicant (for all designated States except US): ZENECA LIMITED [GB/GB]; 15 Stanhope Gate, London W1Y 6LN

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(72) Inventors; and

(75) Inventors/Applicants (for US only): FRAY, Rupert, George [GB/GB]; University of Nottingham, School of Biological Sciences, Division of Plant Science, Sutton Bonington Campus, Loughborough LE12 5RD (GB). THROUP, John, Peter [GB/US]; SmithKline Beecham, Pharmaceuticals R & D, UP1345 South Collegeville Road, P.O. Box 5089, Collegeville, PA 19426 (US). WALLACE, Andrew, David (81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

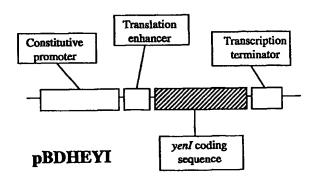
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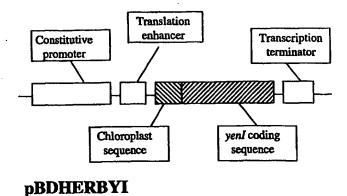
With international search report.

#### (54) Title: EXPRESSION OF BACTERIAL SIGNAL MOLECULES IN PLANTS

#### (57) Abstract

The ability of a plant to defend against attack by bacteria, and any virus borne by the bacteria, is enhanced by transforming the plant genome with a gene of bacterial origin which enables the plant to produce a bacterial pheromone, N-acyl-L-homoserine lactone. Such plants also secrete the lactone into the soil enhancing the protective effect of antifungal rhozobacteria.





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#### **EXPRESSION OF BACTERIAL SIGNAL MOLECULES IN PLANTS**

This invention relates to the expression of bacterial signal molecules in plants which allows, for example, modulation of the interaction between plants and infecting or symbiotic bacteria.

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The ability of bacteria to respond to environmental cues such as nutrient availability, temperature or pH is critical to microbe success. It is apparent that individual bacteria can also sense the density and state of the local bacterial population of which they are members. This sensing ability, referred to as "quorum sensing", allows a bacterial community to synchronise growth and development and, when the minimum population or "quorum" has been achieved, to initiate a concerted population response. Quorum sensing is thus an example of multicellular behaviour in prokaryotes and regulates diverse physiological processes including bioluminescence, swarming, antibiotic biosynthesis, plasmid conjugal transfer and the production of virulence determinants in pathogens.

The signalling pheromones upon which quorum sensing is based have been identified as N-acyl-L-homoserine lactones (reviewed by Swift et.al. "Quorum sensing: a population-density component in the determination of bacterial phenotype", Trends in Biochemical Science, 21, 214-219 (1996). N-acyl-L-homoserine lactones molecules comprise a homoserine lactone moiety (derived from amino acid metabolism, possibly via S-adenosyl methionine) linked to an acyl sidechain (probably derived from fatty acid synthesis). A number of N-acyl-L-homoserine lactones with different acyl side chains have been identified in different bacterial systems where they elicit a wide range of quorum-dependent responses such as swarming, pathogenicity, conjugation or production of colour, light or antibiotics.

Several bacterial species produce the same N-acyl-L-homoserine lactone, although in some of the species it may regulate a different biological process. For example, the luxI gene product of Photobacterium fischeri synthesises N-(3-oxohexanoyl)-L-homoserine lactone which regulates bioluminescence in a cell density-dependent manner, whilst the carI gene product of Erwinia carotovora also produces N-(3-oxohexanoyl)-L-homoserine lactone which in this bacterium is responsible for the induction of secreted plant cell wall degrading exoenzymes and of the antibiotic carbapenem. The cviI gene of Chromobacterium violaceum encodes the enzyme for synthesis of N-hexanoyl-L-homoserine lactone which is

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structurally very similar to the oxohexanoyl analogue and which induces production of the purple pigment violacein. Inactivation of luxI, carI or cviI results in loss of the density dependent bioluminescence, virulence or violacein production respectively. The relevant operons can, however, be induced by the addition of an exogenous supply of the N-acyl-Lhomoserine lactone to the mutant bacteria.

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Carl mutants of Erwinia carotovora appear to be completely avirulent when tested on tobacco. They can neither macerate plant tissue nor multiply in planta because they lack pectin lyase, pectate lyase, polygalacturonase, cellulase and protease. It is pertinent to ask how the expression of these exoenzymes only at high cell density in the wild-type cells may contribute to the success of Erwinia as a plant pathogen. It has been suggested that under aerobic conditions, a successful E.carotovora infection requires a relatively high inoculum (10<sup>6</sup> - 10<sup>7</sup> c.f.u.) and the progression of the disease is then a competition between bacterial multiplication and development of plant resistance. Thus, the production of macerating enzymes at low cell densities would not give rise to a successful infection, but would result in the induction of the local and systemic plant defence response, which in turn would hamper subsequent infections. Such resistance to E.carotovora infection is seen when the plant defence response is artificially induced by the application of salicylic acid.

While not wishing to be bound by any theory as to the manner in which the invention proposed herein operates, the following explanation of the naturally occurring phenomenon of quorum sensing is offered. Using Photobacterium fischeri as a convenient example, the expression of two regulatory genes, luxI and luxR, is necessary for the expression of the genes necessary for bioluminescence. Expression of lux I leads to production of the pheromone N-(3hydroxyl)hexanoyl-L-homoserine lactone, the mechanisms by which the lactone is synthesised being largely irrelevant to this discussion. A complex of the pheromone with the protein produced by the lux R gene gives a phenotypic response, in the case of P. fischeri, bioluminescence. At low population density of bacteria, lux I and lux R are transcribed at low level and there is insufficient accumulation of the pheromone (N-acyl-L-homoserine lactone) to elicit luxI-dependent transcription of the operon responsible for visible bioluminescence. It has been suggested that in the absence of sufficient pheromone, and/or a chaperonin known as GroESL, luxR is unstable and sensitive to degradation. As the population grows, however, the concentration of the pheromone increases gradually. At a critical level of the pheromone, which represents a critical population density, a complex between luxR and the pheromone is thought to bind to a palindromic sequence within the luxI

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operator thereby activating increased transcription of the operon necessary for increased production of the pheromone and for bioluminescence.

The present invention seeks to provide a method and means of manipulating plant/microbe interactions.

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According to the present invention there is provided a method of protecting a plant against bacterial infection and/or virus infection transmitted by bacteria, comprising introducing into the genome of the plant by transformation the ability to synthesise a N-acyl-L-homoserine lactone.

Further according to the invention there is provided a method of protecting a plant against bacterial infection and/or virus infection transmitted by bacteria, comprising introducing into the genome of the plant by transformation the ability to synthesise an analogue of N-acyl-L-homoserine lactone.

The invention also provides a method of enhancing interaction between an antifungal rhizobacterium and a plant comprising introducing into the genome of the plant by transformation the ability to synthesise the *N*-acyl-L-homoserine lactone naturally produced by the rhizobacterium.

The invention also provides a recombinant plant genome containing a gene construct for *in planta* expression of an *N*-acyl-L-homoserine lactone.

Preferably expression of introduced genes is targeted to plant chloroplasts.

The gene specifying the N-acyl-L-homoserine lactone may be selected from the group consisting of, the yenI gene of Yersinia enterocolitica; the cviI gene of Chromobacterium violaceum; the luxI gene of Photobacterium fischeri; the carI gene of Erwinia carotovora; the traI gene of Agrobacterium tumefaciens and the lasI and vsmI genes of Pseudomonas aeruginosa.

Examples of suitable sources of DNAs specifying N-acyl-L-homoserine lactones and the acyl group involved are as follows:

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Table 1

N-acyl-L-homoserine lactone

Bacterium	Signal	Response	N-acyl-group
	Generator	Regulator	R
Yersinia enterocolitica	yenI	yenR	3-oxohexanoyl
Chromobacterium violaceum	cvil	cviR	3-hexanoyl
Photobacterium fischeri	luxI	luxR	3-oxohexanoyl
Erwinia carotovora	carl	carR	3-oxohexanoyl
Agrobacterium tumefaciens	traI	traR	3-oxo-octanoyl
Pseudomonas aeruginosa	lasI	lasR	3-oxo-dodecanoyl
Pseudomonas aeruginosa	vsmI	vsmR	butanoyl

These examples in Table 1 are quoted in Swift et.al., Trends in Biochemical Science, 21, 214-219 (1996).

Table 2 below gives further examples along with references and the appropriate GenBank Accession Numbers.

Table 2

Organism	Signal generator	Response Regulator	Signal Molecule	GenBank Accession number	References
Aeromonas hydrophila	AhyL	AhyR	unknown	X89469	
Agrobacterium tumefaciens	Tral	TraR	N-(3-oxo)- octanoyl-L- homoserine Lactone (OOHL)	L17024, L22207	Fuqua et.al, 1994; Hwang et.al. 1995
Chromobacterium violaceum	CviI	CviR	N-hexanoyl-L-homoserine lactone (OHL)		Winson,et.al, (1994)
Enterobacter	Eagl	unkn wn	N-(-3-	x74300	Swift et al., 1993

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			N	· · · · · · · · · · · · · · · · · · ·	
agglomerans			oxo)hexanoyl-L-		
			homoserine		
			lactone (OHHL)		
Erwinia	Carl	CarR	OHHL	U17224,	McGowan et.al.,
carotovora subsp				X72891,	1995
carotova				X74299,	
				X80475	
Erwinia stewartii	Esal	EsaR	OHHL	L32183,	Beck von
				L32184	Bodman and
					Farrand, 1995
Escherichia coli	unknown	SdiA	unknown	Xo3691	Sitnikov et al
					1995
Photobacterium	Luxl	LuxR	OHHL,OOHL	M19039,	Meignhen, 1994;
fischeri		Ì		M96844,	Devine et al,
				M25752	1988
Pseudomonas	Lasi	LasR	N-(-3-oxo)-	M59425	Winson et al
aeruginosa			dodecanoyl-L-		1995;.
3			homoserine		
			lactone (OdDHL)		
	Vsml	vsmR	N-butanoyl-L-	L08962,	Winson et al.,
	7 2		homoserine	U11811,	1995; Latifi et al
			lactone (BHL),	U15644	1995; Ochsner
			HHL	[	and Reiser, 1995.
Pseudomonas	Phzi	PhzR	unknown	L32729,	Wood and
aureofaciens				L33724	Piersen, 1996
Rhizobium	unknown	RhiR	N(-3-hydroxy)-	M98835	Fuqua et al.,
leguminosarum			tetradecanoul-L-		1994; Gray et al.,
	ļ		homoserine	]	1996.
		ļ	lactone		
			(HtDeHL)		<u> </u>
Serratia	Swrl	unknown	BHL	U2823	
liquefaciens	J				
Aeromonas	ahyl	ahyR	BHL		Swift et al., 1997
hydrophila					
Aeromonas	Asal	unknown	BHL, N-hexanoyl-		Swift et al., 1997
salmonicida			L-homoserine		
			iactone		
Vibiro	vanI	vanR	N-(3-oxo-	†	Milton et al.,

anguillarum			decan yl)-L-		1997
			homoserine		
			lactone (ODHL)		
Vibrio harveyi	LuxLM	LuxN	N-(3-hydroxy)-	L13940	Meighen, 1994;
			butanoyl-L-		Bassler et al.,
		:	homoserine		1994.
		•	lactone (HBHL)		
Yersinia	YenI	YenR	OHHL,HHL	X76082	Throup et al.,
enterocolitica				ŀ	1996.

References:

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Bassler et.al. Molecular Microbiology, 12, 403-412 (1994)

Beck et.al. J.Bacteriol., 177, 5000-5008 (1995) Devine et.al. Biochemistry, 27,837-842 (1988) Fuqua et.al. J.Bacteriol. 176, 269-275 (1994) Gray et.al. J.Bacteriol. 178, 372-376 (1996) Hwang et.al. J.Biotech. 177, 449-458 (1995)

Latifi et.al. *Molecular Microbiology*, 17, 333-343(1995) McGowan et.al. *Microbiology*, 141, 541-550 (1995)

Meignhen Ann. Rev. Genet. 28,117-139(1994)
Milton et.al. J. Bacteriol. 179, 3004-3012 (1994)

Ochsner and Reiser Proc. Natl. Acad. Sci. USA, 92, 6424-6428 (1995)

Sitnikov et.al. *Molecular Microbiology*, **17**, 801-812 (1995 Swift et.al. *Molecular Microbiology*, **10**, 511-520 (1993)

Swift et.al. J. Bacteriol. 179, 5271-5281 (1997)

Throup et.al. Molecular Microbiology, 17, 345-356 (1996) Winson et.al. Proc. Natl. Acad. Sci USA, 92, 9427-9431 (1995)

Wood and Piersen, Gene 168, 49-53 (1996)

Our invention is founded on our reasoning that if the inoculating bacteria were to encounter levels of N-acyl-L-homoserine lactone that gave a false indication of the local population size, the course of the ensuing infection would be drastically reduced.

A second aspect of the invention concerns engineering the plant to take advantage of the potential protective effect of antifungal rhizobacteria. There exist in the rhizosphere certain bacteria which are capable of attacking potential pathogenic fungal microorganisms which are also present in the soil, perhaps the best known of which are certain strains of *Pseudomonas fluorescens* and *P. aureofaciens*. But the population of such antifungal bacterial strains in the soil will normally be low and their antifungal activity dependent on the quorum sensing phenomenon to be activated. By imparting to the plant the ability to produce the activator molecule, the *N*-acyl-L-homoserine lactone, appropriate to the antifungal bacteria the antifungal activity may be initiated at low colony size providing earlier than normal protection of the plant against the pathogenic fungi. The rhizosphere-

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expressed genes of the rhiABC operon of the symbiotic nitrogen-fixing bacterium Rhizobium leguminosarum, for example, are regulated by an AHL with a C14 side chain containing hydroxylated carbon in the 3 position and a single carbon-carbon double bond.

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Transgenic plants producing an AHL signal molecule enhance the establishment of an antifungal environment on the rhizosphere. This phenomenon would also enable the use of disarmed bacterial strains to be used as crop protection biocontrol agents in conjunction with the AHL-producing transgenic plants.

The invention will now be described in the following Examples The ability of AHLs to induce changes in neighbouring bacteria was tested in four

- (1) the ability of the AHLs to diffuse out of intact leaves was demonstrated by placing intact transgenic leaves on agar and subsequently removing it before overlaying with C. violaceum CV026 (see Example 4 below) and the outline of the whole leaf could be seen showing that the AHL diffused out of the leaf surface and not just the cut stem;
- (2) being interested in whether the AHLs were only produced in the chloroplasts or whether they could be found in other tissues such as roots, the ability of the AHLs to diffuse from the roots was demonstrated in that AHLs in the vicinity of the roots were able to induce bioluminescence in a recombinant E, coli strain carrying an AFIL-inducible operon: this showed that the root plastids are competent to suynthesise the AHLs are, alternatively, that the AHLs synthesised in green tissue can be transported to the roots but in either case the roots were clearly capable of signalling to nearby bacteria.
- (3) AHL-producing plant tissue is capable of restoring G.graminis growth-inhibiting activity to the disarmed P. aureofaciens 30-84 phzI- strain (see Example 9 below)
- (4) Erwinia carotovora carI (expI) mutants, which have greatly reduced virulence in their natural host plants were shown to infect transgenic tobacco plants which are producing AHLs (see Example 10 below).

Figure 1 herewith shows the components of the constructs pBDHEYI and pBDHERBYI described in the Examples.

#### Example 1

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#### Preparati n fpBDHEYI

pBDHEYI was constructed by fusing the alfalfa mosaic virus (AMV) translation enhancer sequence from pBI526 (Datla et.al., Plant Science 94, 139-149 (1993)) to the yenI coding sequence from Yersinia enterocolitica. The yenI sequence had previously been amplified by PCR to create an NcoI site overlapping the translation initiation sequence. This changed the second amino acid from leucine to valine but did not affect the ability of the encoded enzyme to synthesise N-acyl-L-homoserine lactones in a bacterial assay. The AMV/yenI fusion was cloned on a Bg/II/BamHI fragment into the BamHI of pDH51 (Pietrzak et.al., Nucl. Acids Res. 14, 5857-5868(1986)) to give pDHEYI. An EcoRI fragment of pDHEYI was cloned into the EcoRI site of pBIN19 (Bevan, Nucl. Acids Res. 12, 8711-8721 (1984)) to give pBDHEYI.

#### Example 2

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#### Preparation of pBDHERBYI

pBDHERBYI was constructed by fusing the petunia SSU611 ribulose bisphosphate carboxylase small subunit (rbcS) chloroplast targeting sequence (Dean et.al. Mol, Gen. Genet., 206, 465-474 (1987)) to the AMV translation enhancer sequence of pBI526. An NcoI site was engineered to overlap the initiating ATG codon of rbcS. An SphI site was engineered to overlap the initiating ATG codon of yenI and the yenI coding sequence cloned into the SphI site of the SSU611 fragment. This site spans the cleavage site of the encoded chloroplast transit peptide. The AMV/rbcS/yenI fusion was cloned on a BglII/BamHI fragment into the BamHI site of pDH51 to give pDHERBYI. An EcoRI fragment from pDHERBYI was cloned into the EcoRI site of pBIN19 to give pBDHERBYI.

The rationale for producing pBDHERBYI and believing that it would be active in chloroplasts was as follows: in *E.coli* homoserine lactone is not produced by mutants of the threonine biosynthetic pathway that are blocked prior to homoserine synthesis but is produced by those mutants when supplied with an exogenous source of homoserine. However, *TraI*, the *N*-acyl-L-homoserine lactone biosynthetic enzyme in *Agrobacterium tumefaciens*, has been found to utilise *S*-adenosylmethionine and not homoserine as a substrate *in vitro*. There is also evidence for the acyl moiety being derived from fatty acid biosynthetic intermediates. In plants the enzymes of the threonine biosynthetic pathway are located in the chloroplast and this organelle is also active in fatty acid metabolism. Therefore the chloroplasts may be expected to contain the necessary precursors for *N*-acyl-L-homoserine lactone synthesis by *yenI* and more closely approximate to the environment in which *yenI* is normally active than would be the cytoplasm.

#### Example 3

Generation of Transgenic Plants

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Construct pBDHEYI.for Example 1 and pBDHERBYI from Example 2 were transferred to the *Agrobacterium tumefaciens* strain LBA 4404 and used to transform tobacco leaf discs according to standard protocol (Draper et.al., pages 69-160, In Plant Genetic Transformation and gene expression: a laboratory manual; Draper et.al. (Eds) Blackwell Scientific Publications, London (1988)).

The transgenic status of the resulting kanamycin positive explants was confirmed by Southern analysis (data not given)

#### Example 4

#### Complementation of Violacein Production

Leaf segments of the transgenic plants produced in Example 3 were tested for their ability to synthesise N(3-oxohexanoyl)-L-homoserine lactone or a related analogue.

A transgenic tobacco leaf was placed in an agar plate overnight. The leaf was then removed and the *cviI* mutant of *Chromobacterium violaceum* spread over the plate. Violacein production by the bacteria could be seen where the *N*-(3-oxohexanoyl)-L-homoserine lactone had diffused out of the leaf and into the agar.

Two leaf segments tested positive as indicated by the ability of a diffusible product to complement *C.violaceum*, inducing the production of the purple pigment violacein by the bacteria.

#### Example 5

#### Complementation of carI

Construct pBDHERBYI (Example 2) was transferred to the Agrobacterium tumefaciens strain LBA 4404 and transformed into tobacco. Leaf segments were tested for their ability to synthesise N(3-oxohexanoyl)-L-homoserine lactone or a related analogue.

An untransformed control and a transgenic BDHERBYI tobacco leaf were inoculated with *Erwinia carotovora* mutant for *carI*. The bacteria were applied at a high culture density (OD600 of 2.5) in a volume of 10 µl to a small wound site made with a hypodermic needle. A second BDHERBYI leaf was mock inoculated with bacterial culture medium alone.

The leaves were inspected after four days. The untransformed control and the mock inoculated leaf remained substantially unchanged. The sample inoculated with *E.carotovora* displayed advanced disease symptoms demonstrating that the pathogen can perceive and respond to the *N*-acyl-L-homoserin lactone being made by the transgenic plant.

#### Example 6

#### Complementati n fluxI

Following a similar protocol as described above, the *luxR N*-acyl-L-homoserine lactone response regulator and the *lux* operon (minus *luxl*) of *Pseudomonas fischeri* was inserted into *E.coli*. When transgenic tobacco carrying the BDHERBYI construct was challenged with the *E.coli*, bioluminescence was induced in the bacteria demonstrating that the *luxR* gene was able to respond to the *N*-acyl-L-homoserine lactone produced by the plant.

Twenty-nine tobacco plants that were independently transformed with either BDHERBYI or BDHEYI were challenged with *C.violaceum* mutant for *cviI* (Example 3) and *E.coli* carrying an *N*-acyl-L-homoserine lactone-inducible *lux* operon. Table I summarises the results.

	Positive reaction Negative reaction				
Construct	cviI	luxI	cviI	luxI	Total
BDHERBYI	8	8	5	5	13
BDHEYI	0	0	16	16	16

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# Example 7 Extraction and TLC analysis of AHLs

For thin-layer chromatographic analysis, transgenic plant extracts were made by grinding two grams of plant tissue to a fine powder in liquid nitrogen and mixing the frozen powder with 200ml of warn distilled water. After five minutes, solid matter was filtered off and the filtrate extracted with an equal volume of ethyl acetate. The ethyl acetate layer was then dried over anhydrous magnesium sulphate, filtered and evaporated to dryness. The residue was taken up in 500 µl of acetonitrile and 20µl of this was applied to a C18 reverse phase TLC plate (Merck). A similar extract from an untranformed control plant was also spotted on to the plate. N-hexanoyl-L-homoserine lactone (HHL) (1x10-8g) and N-(3-oxohexanoyl) -L homoserine lactone (OHHL) (5x10-7g) were applied as standards and the chromatogram developed with methanol/water (60:40 vol/vol) as running solvent (Shaw P.D. et.al. Proc.Natl.Acad,Sci. USA 94: 6036-6041 (1997)). After drying, AHLs were located on the TLC plate by overlaying C.violaceum strain CV026 in top agar as described by McClean et.al. (Microbiology-UK, 143: 3703-3711 (1997). After 16 hours growth at 28°C the presence of AHLs was indicated by localised violacein production.

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Two different molecules with  $R_f$  values identical to the synthetic HHL and OHHL standards were observed.

#### Example 8

#### **HPLC and LC-MS Analyses**

For HPLC and LC-MS analyses, transgenic plant extracts were made by grinding the issue in ethyl acetate. The supernatant was taken and the plant residue re-extracted with ethyl acetate, the supernatants pooled and the process repeated until the plant residue was white/brown n colour and free of chlorophyll. The ethyl acetate layer was separated from a small plant-derived aqueous layer and dried over anhydrous magnesium sulphate, filtered, and evaporated to dryness. The residue was resuspended in 500 µl of methanol, this was brought to 60% methanol with sterile distilled water and placed at -20°C overnight to precipitate out the majority of the chlorophyll. After pelleting any solid matter by centrifugation in a bench-top microfuge, the AHL-containing supernatant was partitioned against 10 volumes of ethyla acetate and the organic phase evaporated to dryness. The residue was taken up into 500µl of acetonitrile. For both LC-MS and HPLC analyses linear gradients of acetonitrile in water were run (20-100% over 32 minutes) as described by Camara et.al. In Methods in Microbiology: Bacterial Pathogenesis Vol. 27: 319-330, Williams et.al. (Eds) (1998). OHHL and HHL eluted at 9 minutes and 13.5 minutes respectively.

The presence of HHL and OHHL, detected in TLC analysis, were confirmed.

#### Example 9

#### Assay for restoration of activity to P.aureofaciens mutant

Leaf material from transgenic and non-transformed control plants were placed in wells cut in a potato dextrose agar plate (Oxoid). *P.aureofaciens* strain 80-84I (*phzI*-) was inoculated adjacent to the wells and the plates incubated for 24 hours at 22°C. The *G.graminis* var. tritici was then introduced on the opposite side of the plate and the whole incubated for a further four days.

The antifungal activity of the *P. aureofaciens phzI*<sup>-</sup> strain against the *G. graminis* was found to have been restored.

#### Example 10

Assay for restoration of virulence to Erwinia carotovora avirulent mutant

Untransformed and control BDHERBYI tobacco leaves were inoculated with the avirulent E.carotovora mutant PNP22 (Bainton et.al., Biochem.Journal, 288: 997-1004)

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(1992) and also Jones *et.al.*, The EMBO Journal, 12: 2477-2482 (1993)) The bacteria were applied at high culture density (OD600=2.5) in a volume of 10µl to a small wound site made with a hypodermic needle.

Normally these *Erwinia* mutants are avirulent in the tobacco system, in which they can neither macerate plant tissue nor multiply *in planta* because they are defective in the production of plant cell-wall-degrading enzymes pectin lyase, pectate lyase, polygalacturonase, cellulase and protease. The regulated expression of plant cell wall-degrading enzymes only at high density in wild-type bacteria may contribute to the success of *Erwinia* as a plant pathogen. Under aerobic conditions, *E. carotovora* infection only occurs when the bacteria has reached sufficiently high population densitiy such that disease progression depends on competition between bacterial multiplication and the plant host defences. Thus the production of macerating enzymes at low cell densities would not give rise to a successful infection, but would result in the premature induction of the local and systemic plant defence response, which in turn would hamper subsequent infection. Thus, if the infecting pathogen were to encounter AHL levels that gave a false indication of the local bacterial population size the course of the ensuing infection will be substantially reduced as the plant is able to mount a successful defence to a weak attack.

Other modifications of the present invention will be apparent to those skilled in the art without departing from the scope of the invention.

#### **CLAIMS**

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- A method of protecting a plant against bacterial infection and/or virus infection transmitted by bacteria, comprising introducing into the genome of the plant by transformation the ability to synthesise a N-acyl-L-homoserine lactone.
- 2. A method of protecting a plant against bacterial infection and/or virus infection transmitted by bacteria, comprising introducing into the genome of the plant by transformation the ability to synthesise an analogue of N-acyl-L-homoserine lactone capable of competing with the N-acyl-L-homoserine lactone secreted by infecting bacteria for N-acyl-L-homoserine lactone receptor sites therein.
  - 3. A method of enhancing interaction between a rhizobacterium and a plant comprising introducing into the genome of the plant by transformation the ability to synthesise the *N*-acyl-L-homoserine lactone naturally produced by the rhizobacterium.
- 4. A method as claimed in any of claims 1 to 3 in which the gene expressing the N-acyl-L-homoserine lactone is selected from the group consisting of, the yenI gene of Yersinia enterocolitica; the cviI gene of Chromobacterium violaceum; the luxI gene of Photobacterium fischeri; the carI gene of Erwinia carotovora; the tral gene of Agrobacterium tumefaciens and the lasI and vsmI genes of Pseudomonas aeruginosa.

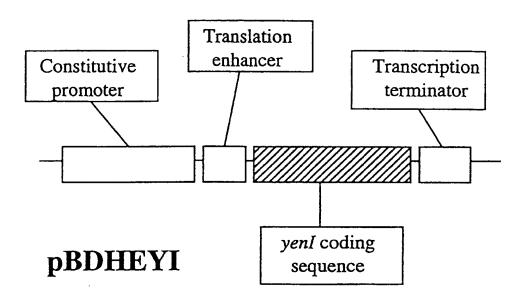
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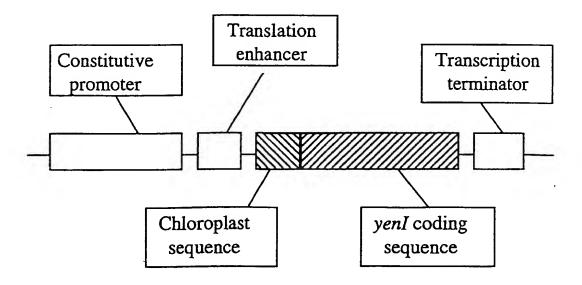
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- 5. A recombinant plant genome containing a gene construct for *in planta* expression of an N-acyl-L-homoserine lactone and/or the response regulator thereof.
- 6. A genome as claimed in claim 5 in which expression of the said N-acyl-Lhomoserine lactone is targeted to plant chloroplasts.

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Figure 1





## **pBDHERBYI**

Intrational Application No Pui/GB 99/02652

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 C12N15/31 C12N15/82 A01H5/00						
According to	International Patent Classification (IPC) or to both national classification	ation and IPC				
B. FIELDS	SEARCHED					
Minimum do IPC 7	cumentation searched (classification system followed by classification CO7K C12N A01H	on symbols)				
Documentat	ion searched other than minimum documentation to the extent that s	uch documents are included in the fletds se	parched			
Electronic d	ata base consulted during the international search (name of data bas	se and, where practical, search terms used				
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT					
Category °	Citation of document, with indication, where appropriate, of the rela	evant passages	Relevant to claim No.			
A	DE 195 48 301 C (DUERING KLAUS DR 27 February 1997 (1997-02-27) the whole document	2)	1-6			
A	WO 97 48719 A (BECKERMAN JANNA L & M UNIVERSITY SYST (US); ZHANG L 24 December 1997 (1997-12-24) the whole document	1–6				
A	WO 96 29392 A (UNISEARCH LTD ;KJE STAFFAN (AU); STEINBERG PETER (AL 26 September 1996 (1996-09-26) the whole document	2				
		-/				
X Furti	her documents are listed in the continuation of box C.	χ Patent family members are listed	in annex.			
° Special ca	tegories of cited documents:					
*Special categories of cited documents:  "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention						
	"E" earlier document but published on or after the international filing date "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to					
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another "Y" document of particular relevance; the claimed invention						
citation or other special reason (as specified)  "O" document referring to an oral disclosure, use, exhibition or  "O" document referring to an oral disclosure, use, exhibition or						
other means ments, such combination being obvious to a person skilled						
aler than the priority date claimed "&" document member of the same patent family						
Date of the	actual completion of the international search	Date of mailing of the international sea	arch report			
2	November 1999	17/11/1999	,			
Name and	mailing address of the ISA	Authorized officer				
	European Patent Office. P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040. Tx. 31 651 epo nl.	Kania, T				
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT  Category Citation of document, with indication, where appropriate, of the relevant passages  Relevant to claim No.						
- ···- y-· ;						
A .	GRAY K. ET AL.: "Cell-to cell signaling in the symbiotic nitrogen-fixing bacterium Rhizobium leguminosarum: autoinduction of a stationary phase and rhizosphere-expressed genes" JOURNAL OF BACTERIOLOGY, vol. 178, no. 2, 1996, pages 372-376, XP002084285 the whole document	3				
A .	ROSEMEYER ET AL: "luxI- and luxR-homologous genes of Rhizobium etli CNPAF512 contribute to synthesis of autoinducer molecules and nodulation of Phaseolus vulgaris" JOURNAL OF BACTERIOLOGY, vol. 180, no. 4, 1 February 1998 (1998-02-01), pages 815-821, XP002084284 ISSN: 0021-9193	3				
Α	THROUP J. ET AL.: MOLECULAR MICROBIOLOGY, vol. 17, 1996, pages 345-56, XP002121181 cited in the application the whole document	1-6				
A	ROBSON N D ET AL: "Bacterial N-acyl-homoserine-lactone-dependent signalling and its potential biotechnological applications" TRENDS IN BIOTECHNOLOGY, vol. 15, no. 11, 1 November 1997 (1997-11-01), page 458-464 XP004092668 ISSN: 0167-7799 see the whole document; esp. p.461 r. col.	1-6				
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Information on patent family members

International Application No Pc./GB 99/02652

Patent document cited in search report		Publication date	Patent family member(s)		Publication date
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